

SCIENTIFIC REPORT

Bupivacaine injection of eye muscles to treat strabismus

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Background: Bupivacaine injected into animal muscles induces a cycle of myotoxicity, degeneration, regeneration and hypertrophy of muscle fibres, without adverse effects on other tissues. This induced hypertrophy can be harnessed to treat strabismus.

Methods: Bupivacaine, 4.5 ml of a 0.75% solution, was injected into the right lateral rectus (RLR) muscle of a patient who had diplopia and who showed 14-prism-dioptres oesotropia.

Results: RLR paresis persisted for 7 days. Then, the RLR regained its abducting ability, and progressive improvement of alignment to 4-prism-dioptres oesophoria occurred over the next 33 days, with the elimination of diplopia. Alignment remained the same at 54 days after injection. Magnetic resonance imaging showed a focal increase in the size of the injected RLR of 58% in the posterior area, with reduced change in anterior portions of the RLR.

Conclusion: Injection of bupivacaine to induce hypertrophy of the injected muscle and thus alter eye alignment was effective in our patient. This approach can be a useful addition to the treatment of strabismus.

Bupivacaine injection into the muscles of laboratory animals has been reported by several authors to show specific myotoxicity for muscle fibres, leaving the basal lamina, nerves and satellite cells intact.^{1–3} Subsequent repair with satellite cell proliferation and eventual muscle hypertrophy then occurs, with some variability depending on the muscle fibre type, fast or slow.^{4–5} Eye muscles are also sometimes injected with bupivacaine during retrobulbar anaesthesia, with the resulting strabismus cases showing enlarged muscles.^{6–10} After appropriate institutional review board procedures, a protocol was established to experimentally test whether bupivacaine injection could be used to alter strabismus in humans in a beneficial way.

CASE REPORT: METHODS AND RESULTS

A woman started having horizontal diplopia at the age of 22 years. Prism glasses treated this successfully for 30 years. At age 56 years, diplopia became a problem, and left medial rectus recession and left lateral rectus resection were performed for 30-prism-dioptres oesotropia. A 4-prism-dioptres residual oesophoria was asymptomatic until age 66 years, with the oesotropia then increasing to 14-prism-dioptres concomitant oesotropia at age 72 years. Vision was 20/30 in the right eye,

reduced by mild cataract formation, and 20/40 in the left eye, reduced by a mild cataract and amblyopia. Fusion and stereopsis were present with prism correction. No evidence of neurological impairment was present, and magnetic resonance imaging of the brain and orbits was normal. Treatment with prism glasses, botulinum toxin injection or surgery was offered. After discussion regarding its experimental nature, informed consent was obtained for an experimental injection. Bupivacaine, 4.5 ml of a 0.75% solution, was injected into the right lateral rectus (RLR) muscle, using the recording of muscle electrical activity from the needle tip to guide the location of the injection. Mild proptosis, a small subconjunctival haemorrhage, diminished abduction of the eye and a slight increase in oesotropia resulted. There was no change in vision and no pain. Abduction weakness persisted for 7 days, followed by recovery of motility and then reduction in the deviation. At 16 days after injection, the primary position deviation was 8-prism-dioptres oesotropia, and diplopia was no longer present. At 33 days after the injection, the deviation was further reduced to 4 prism-dioptres oesophoria. It remained 4-prism-dioptres oesophoria at the most recent examination, 54 days after injection. Magnetic resonance imaging showed an increase in the size of the injected RLR. The cross-sectional area of the RLR in the posterior orbit before injection was 0.382 cm²; at 33 days after injection it was 0.609 cm², an increase of 58%. We used NIH public domain software, ImageJ, to make these measurements and comparisons.¹¹ Reduced hypertrophy was evident in anterior magnetic resonance image slices (fig 1). The cross-sectional area of the other muscles was unchanged.

DISCUSSION

Bupivacaine injection of muscle in laboratory animals results in immediate and massive degeneration of muscle fibres, with dissolution of myofibrils at the Z-band.^{1–2} Other structures are substantially unchanged, including the basal lamina, the satellite cells of which form the muscle fibres, and nearby nerves and vasculature.^{12–15} Inflammatory cells and macrophages remove the degenerated muscle fibres over 2–10 days time.^{2–15} Beginning at about day 2, satellite cells are activated and regeneration begins with the muscle reaching pre-injection size and strength around day 21.^{1–5} The satellite cells continue to elaborate new fibres, with the resulting hypertrophy continuing for many days, and with the muscle remaining enlarged for at least 180 days.^{4–5} The participation of satellite cells in eye muscle function and the signals that control their activation are related fields of great interest.^{16–17} We suppose that the strabismus after cataract surgery also results from muscle hypertrophy induced by bupivacaine.¹⁸ Fibrosis and

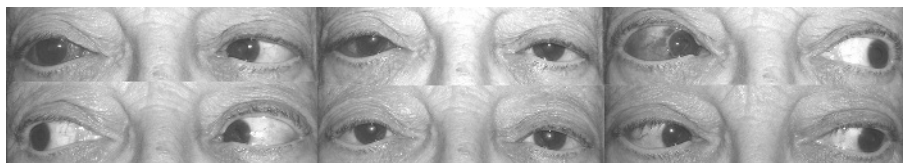


Figure 1 The patient immediately after bupivacaine injection (above), and at 33 days after injection (below). Informed patient consent was obtained for publication of this figure.

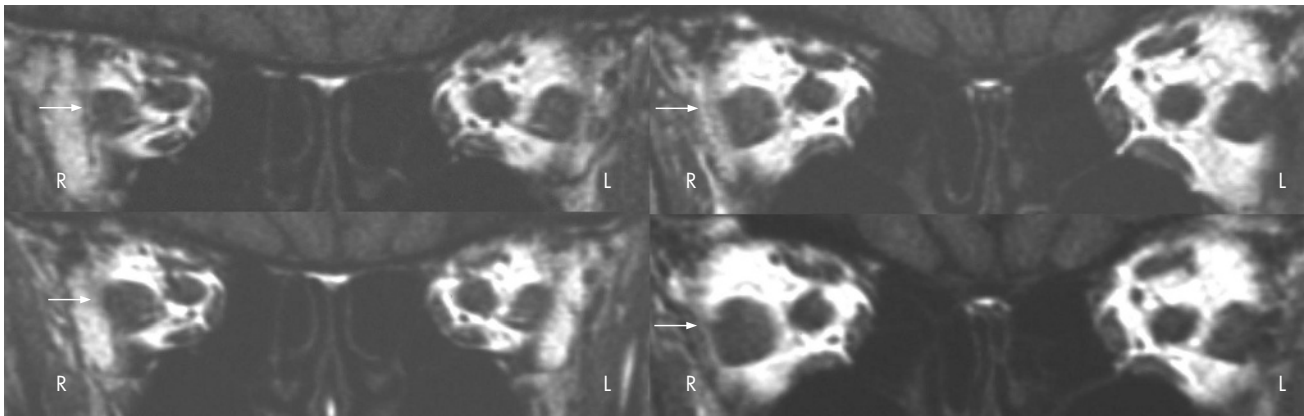


Figure 2 Magnetic resonance imaging T1-weighted coronal scans of the orbits showing images taken before injection (above), and images taken at 33 days after injection (below). The images on the left are at about 12 mm anterior to the orbital apex, and those on the right are at about 16 mm anterior to the orbital apex. The arrows indicate the right lateral rectus muscle. L, left; R, right.

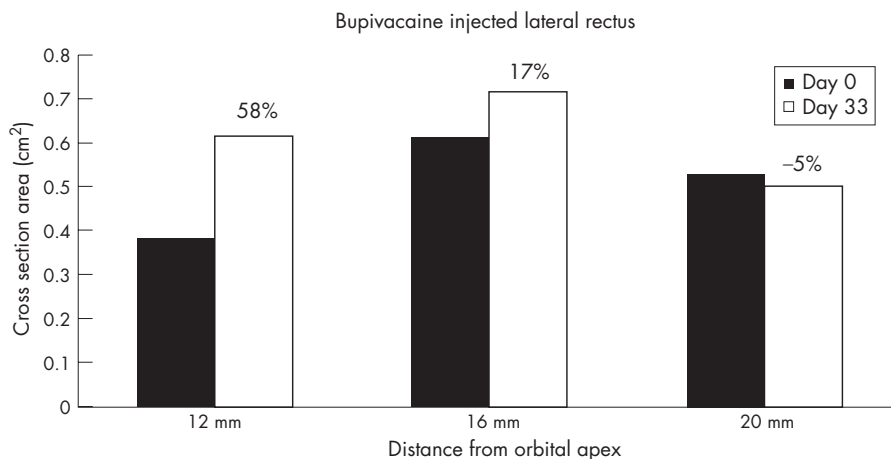


Figure 3 Right lateral rectus cross-sectional areas (cm²) at 12, 16 and 20 mm from the orbital apex, calculated using public domain ImageJ software. Pre-bupivacaine values are shown as solid bars; values at 33 days after injection are shown as open bars. The percentage change is indicated.

scarring, the current putative causes for such strabismus, have not been documented by biopsy, nor found in animal injection studies. Furthermore, the modelling program-Orbit 1.8™ (Eidactics, San Francisco) indicates that muscle hypertrophy, not shortening or stiffening from scarring, can explain the overaction pattern characteristic of these cases.¹⁹ Hypertrophy also occurs after injection of denervated muscle in animals, auguring well for therapeutic trials in paretic strabismus.²⁰ Accurate measurement of eye muscle sizes, forces and changes in alignment are under way in animals and humans in order to define the optimum volume of injection, drug dosage and location in the muscle for injection. It has not escaped our notice that extension of this approach to other muscles holds much promise.

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